

Consensus Sequences: Just Say No!

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<http://www-lmmb.ncifcrf.gov/~toms/>



**CONSENSUS
SEQUENCES**



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Summary

Consensus sequences are being used to characterize the binding sites of macromolecules on DNA and RNA. After aligning a set of binding site sequences, the most frequent base is chosen. A position which contains 100% A's will be represented by an A, while a position that is only 75% A will also be represented by an A. The consensus is frequently used to search for binding sites, and the number of mismatches to the consensus is counted. A mismatch to a 100% A position is much more severe than one to a 75% A, but this is not accounted for so the researcher is misled. We present mathematically robust graphical replacements for the consensus sequence called the sequence logo and the walker that do not discard your hard-earned data. Further information and examples may be found on the internet at <http://www-Immb.ncifcrf.gov/~toms/>.

Consensus Sequences

Characterize what a binding site looks like

❄ Use Sequence Logos instead

Search for new sites

❄ Use Rindividual Matrix Scans instead

Investigate how well the bases of a sequence match to functional binding sites

❄ Use the Walker instead

Summary

Consensus sequences are being used to characterize the binding sites of macromolecules on DNA and RNA. After aligning a set of binding site sequences, the most frequent base is chosen. A position which contains 100% A's will be represented by an A, while a position that is only 75% A will also be represented by an A. The consensus is frequently used to search for binding sites, and the number of mismatches to the consensus is counted. A mismatch to a 100% A position is much more severe than one to a 75% A, but this is not accounted for so the researcher is misled. We present mathematically robust graphical replacements for the consensus sequence called the sequence logo and the walker that do not discard your hard-earned data. Further information and examples may be found on the internet at <http://www-lmmb.ncifcrf.gov/~toms/>.

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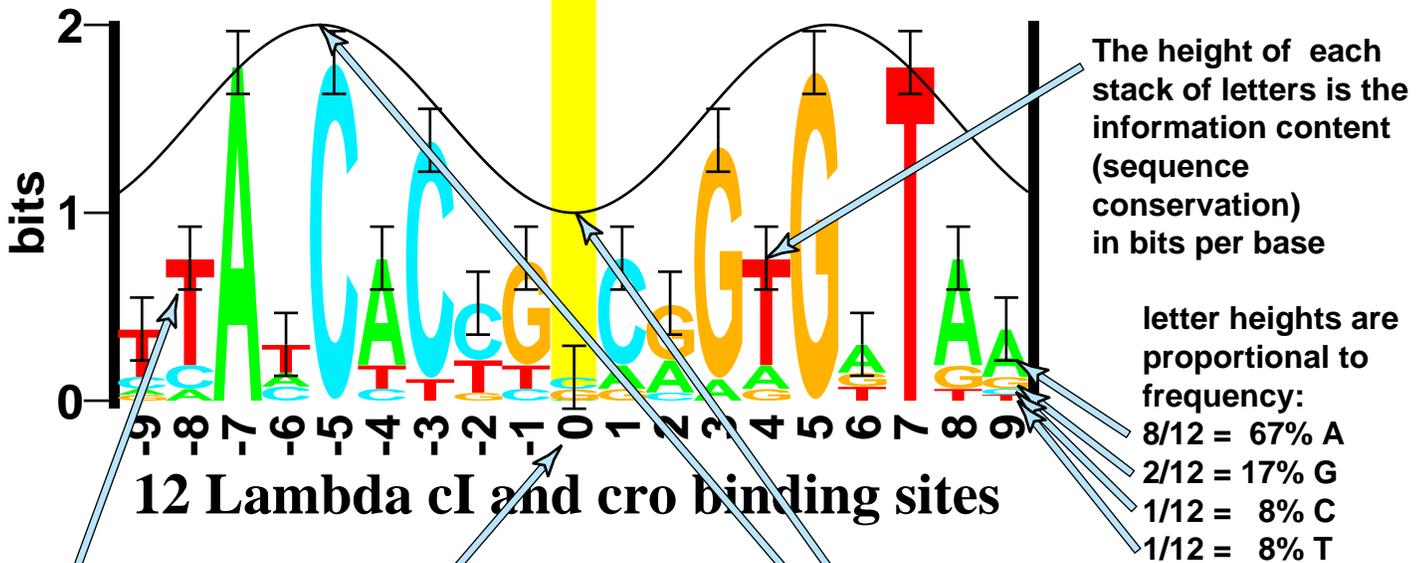
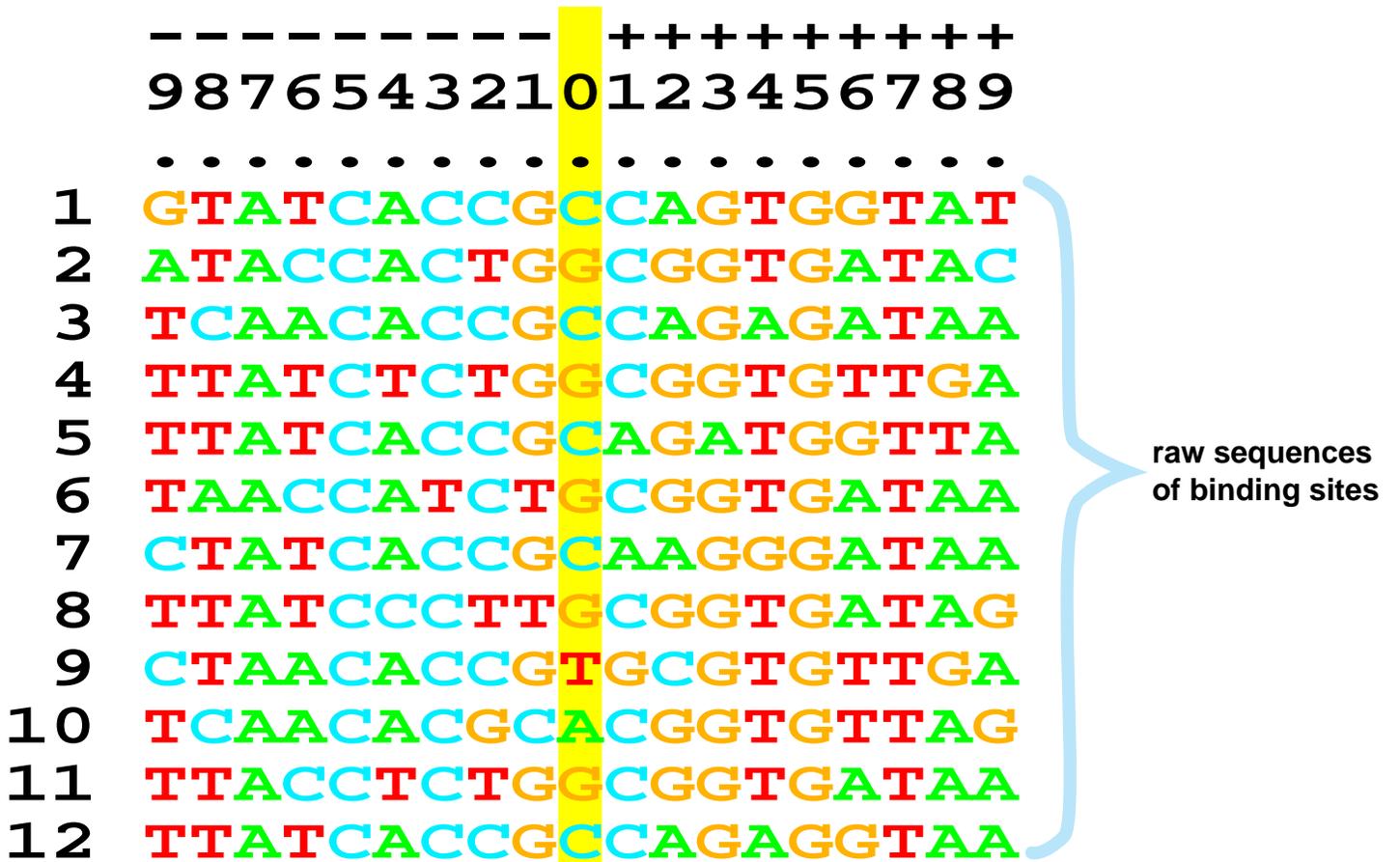
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SEQUENCE LOGO



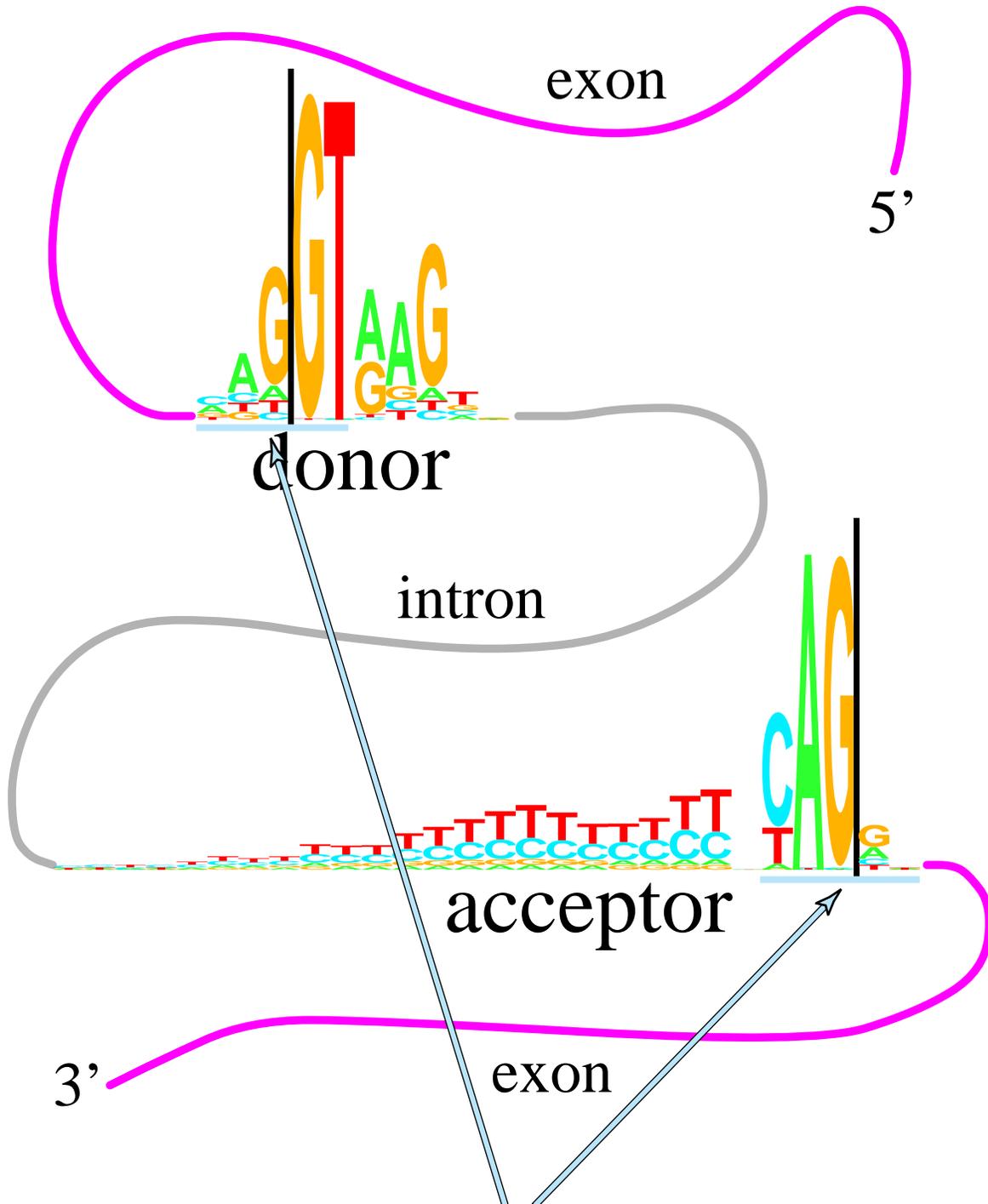
12 Lambda cl and cro binding sites

Error bar for entire stack of letters

Zero coordinate for alignment

Minor Groove Faces Protein
Major Groove Faces Protein

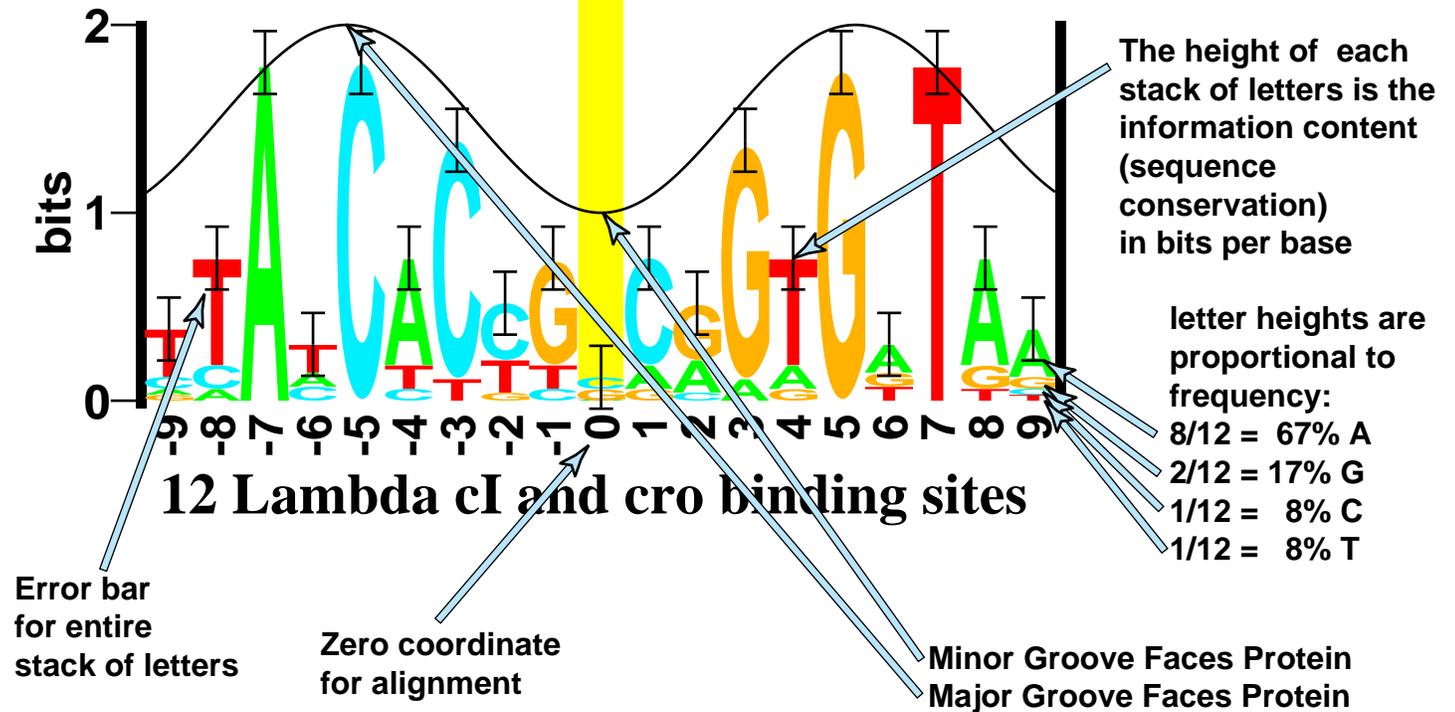
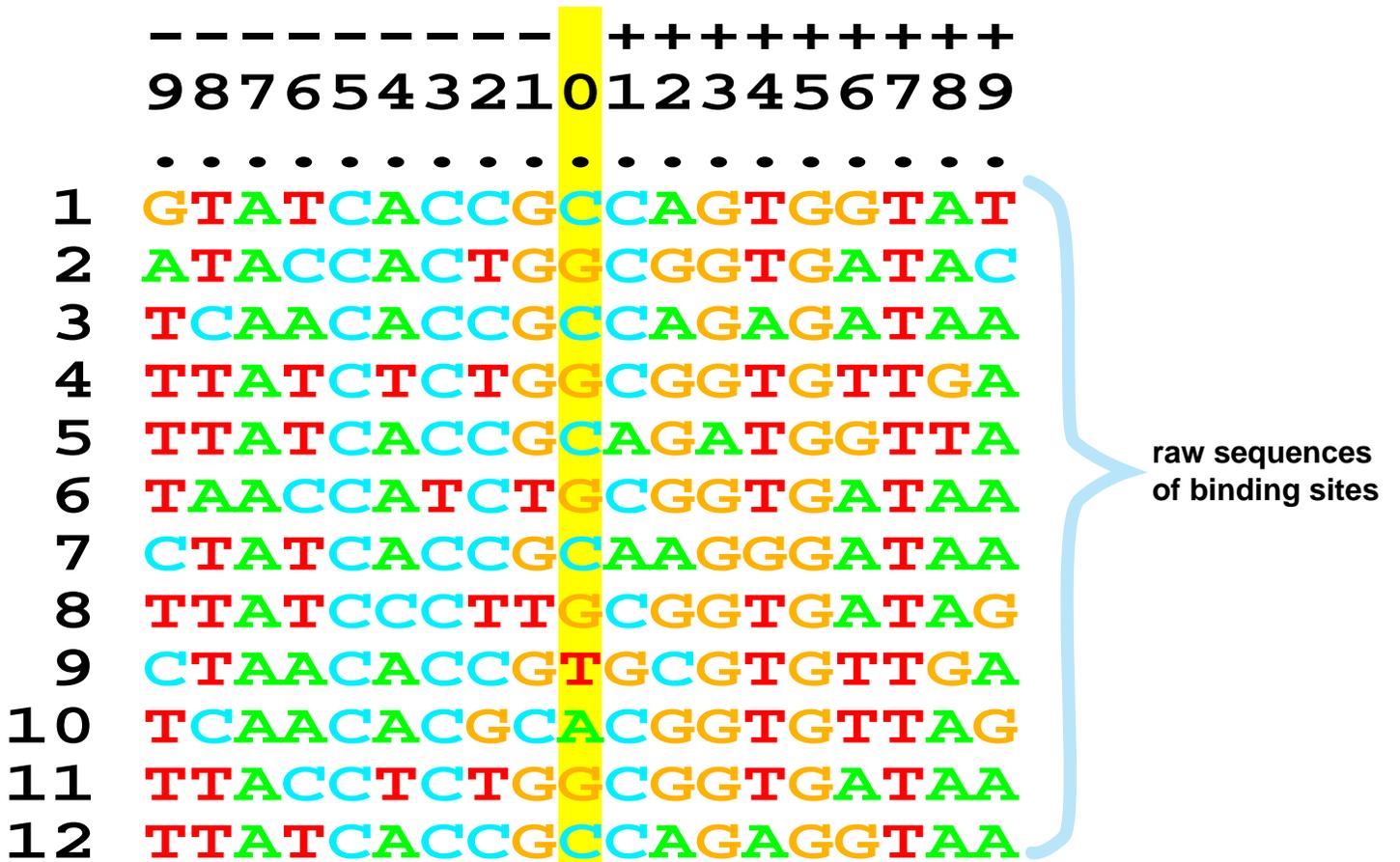
How can two binding sites be different but have the same consensus sequence?



These two sequence logos have the same consensus sequence (CAGGT) but different emphasis



SEQUENCE LOGO

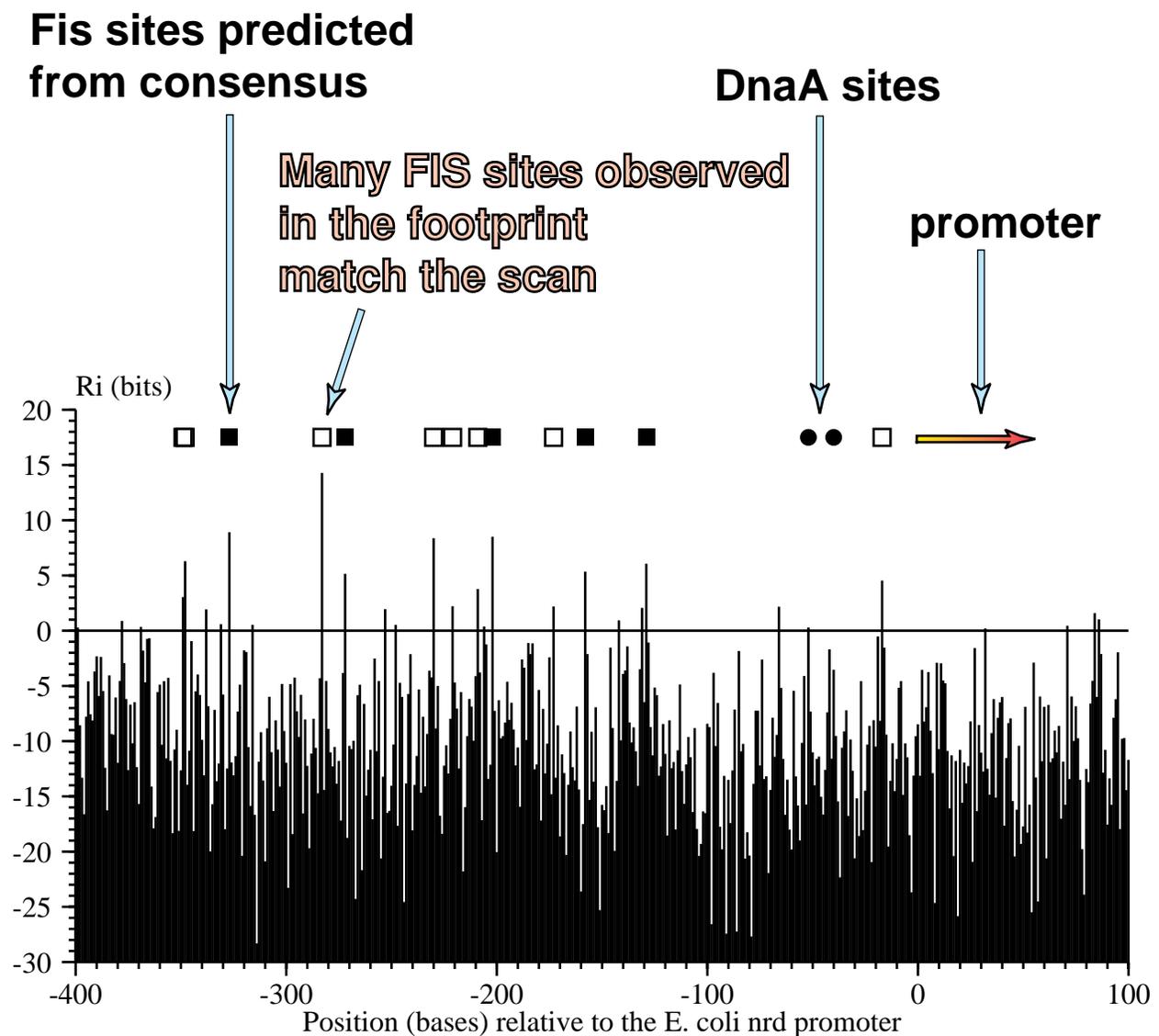


SCAN

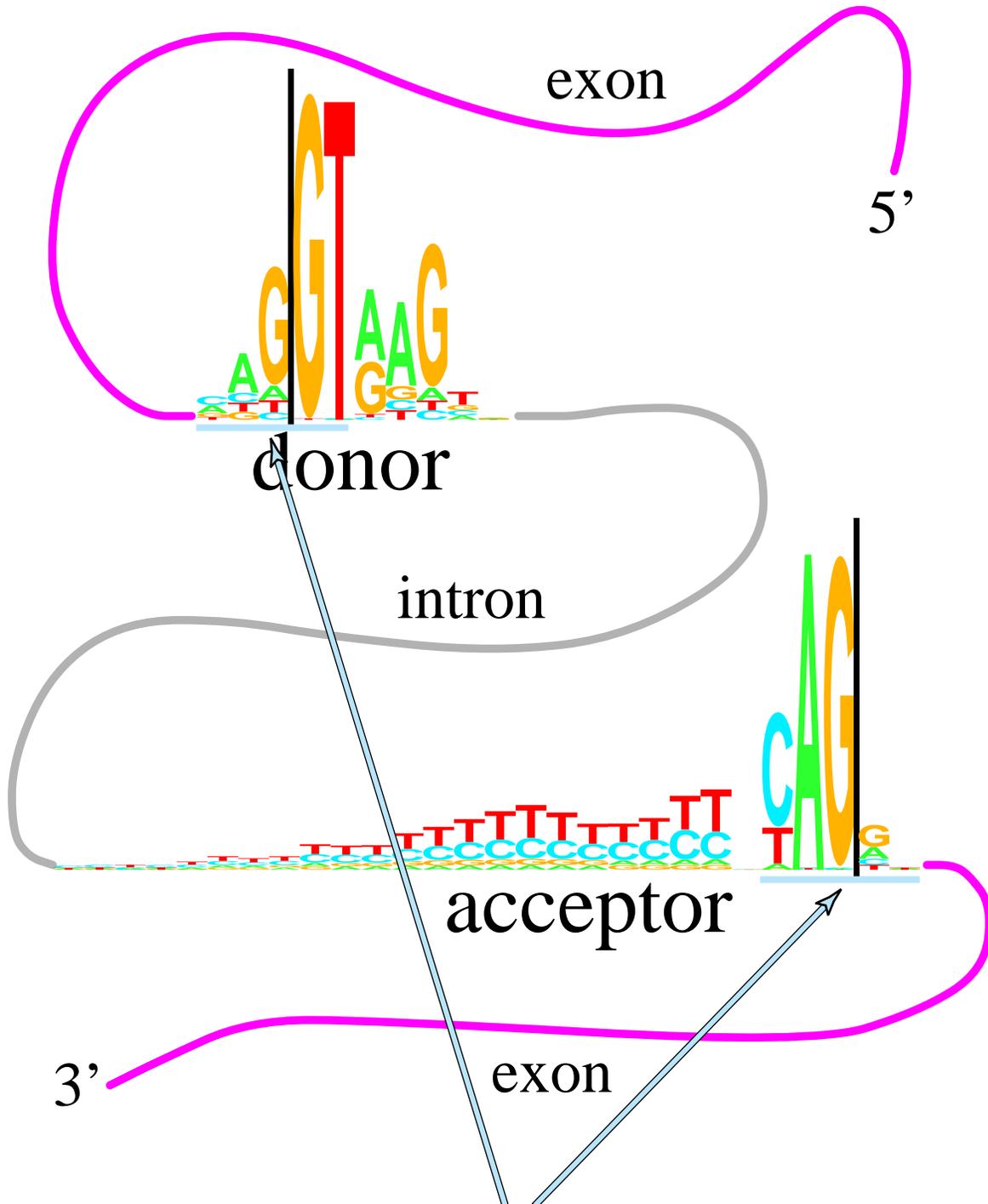
The individual information weight matrix is put at every position of a sequence.

The weights are added together depending on the sequence.

This gives the total Rindividual (Ri) at every position in the sequence.



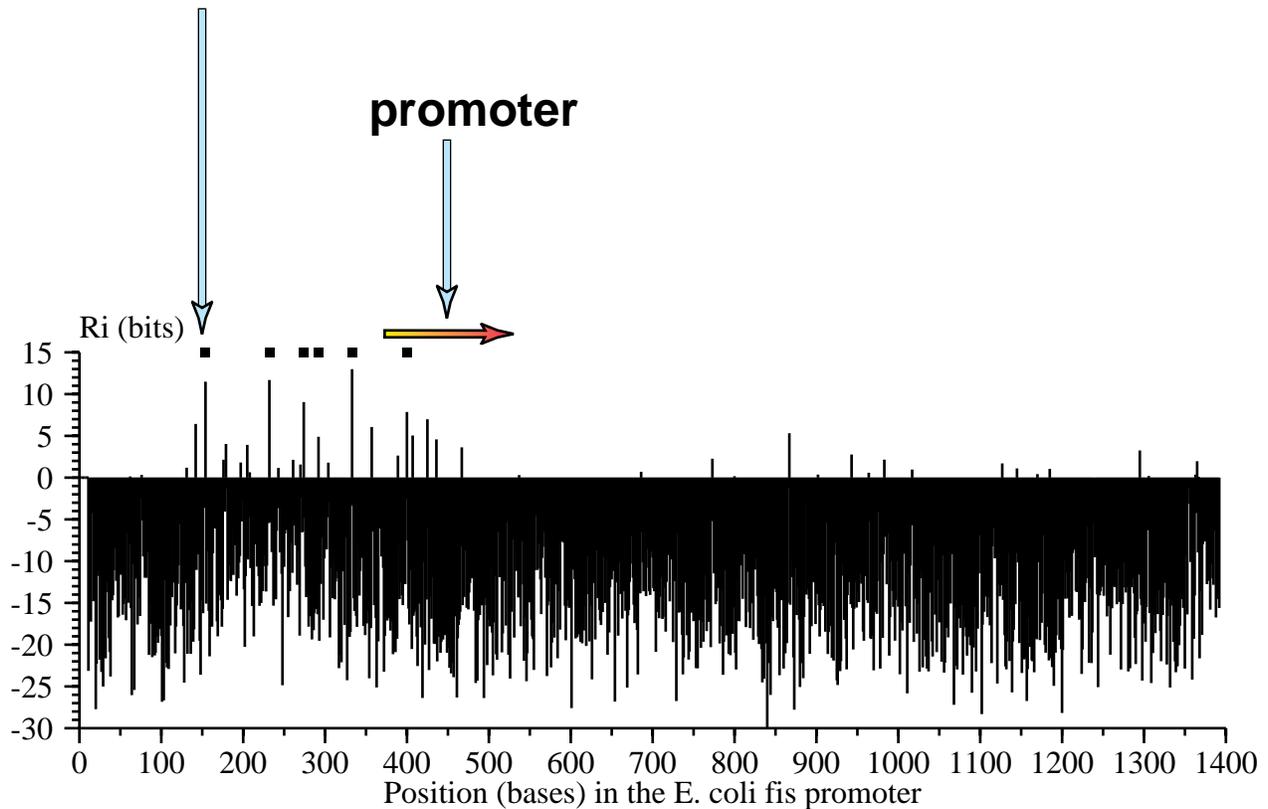
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Scan of Fis Promoter

6 Fis sites were predicted on the *E. coli* Fis promoter from footprints and consensus



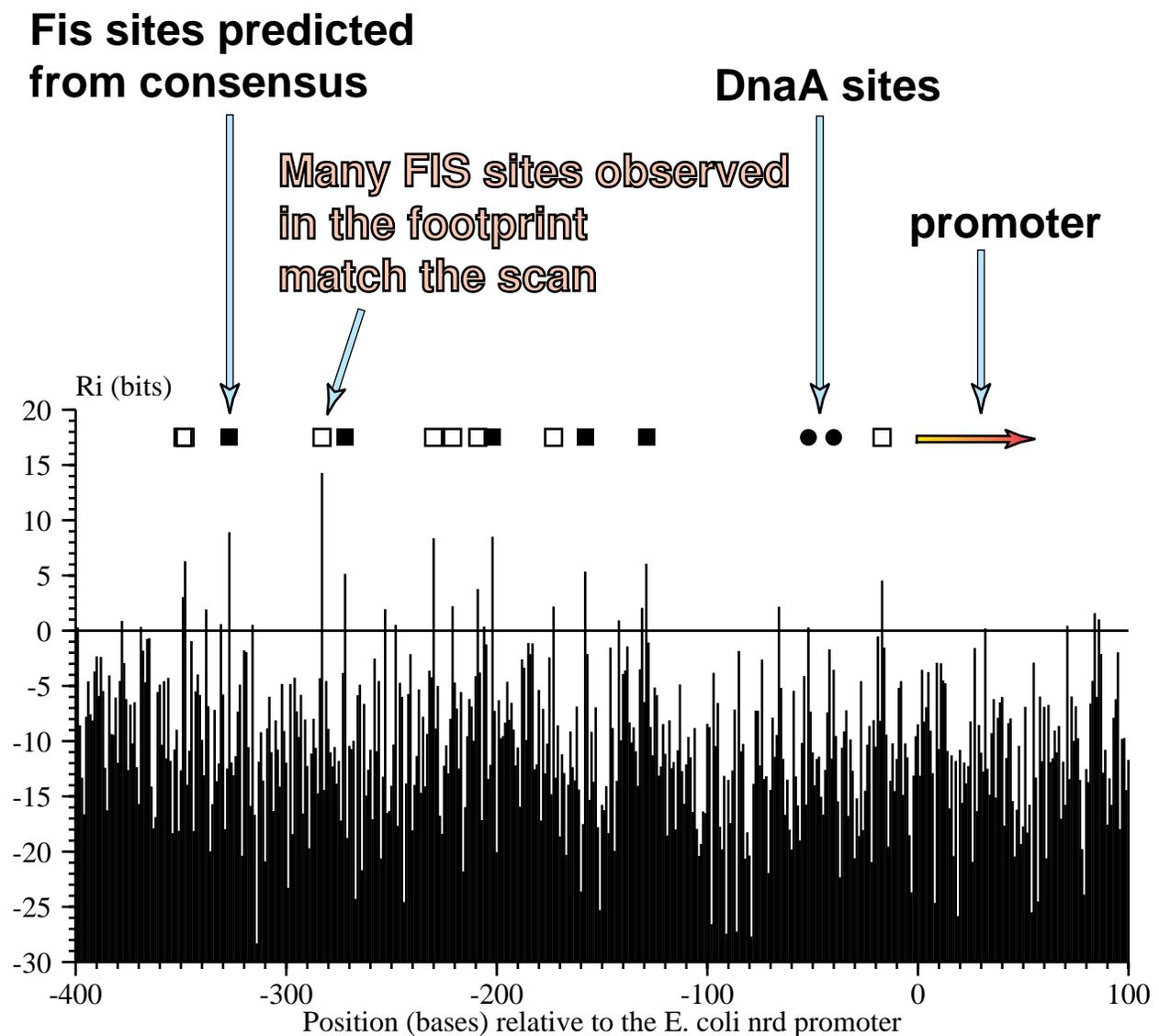
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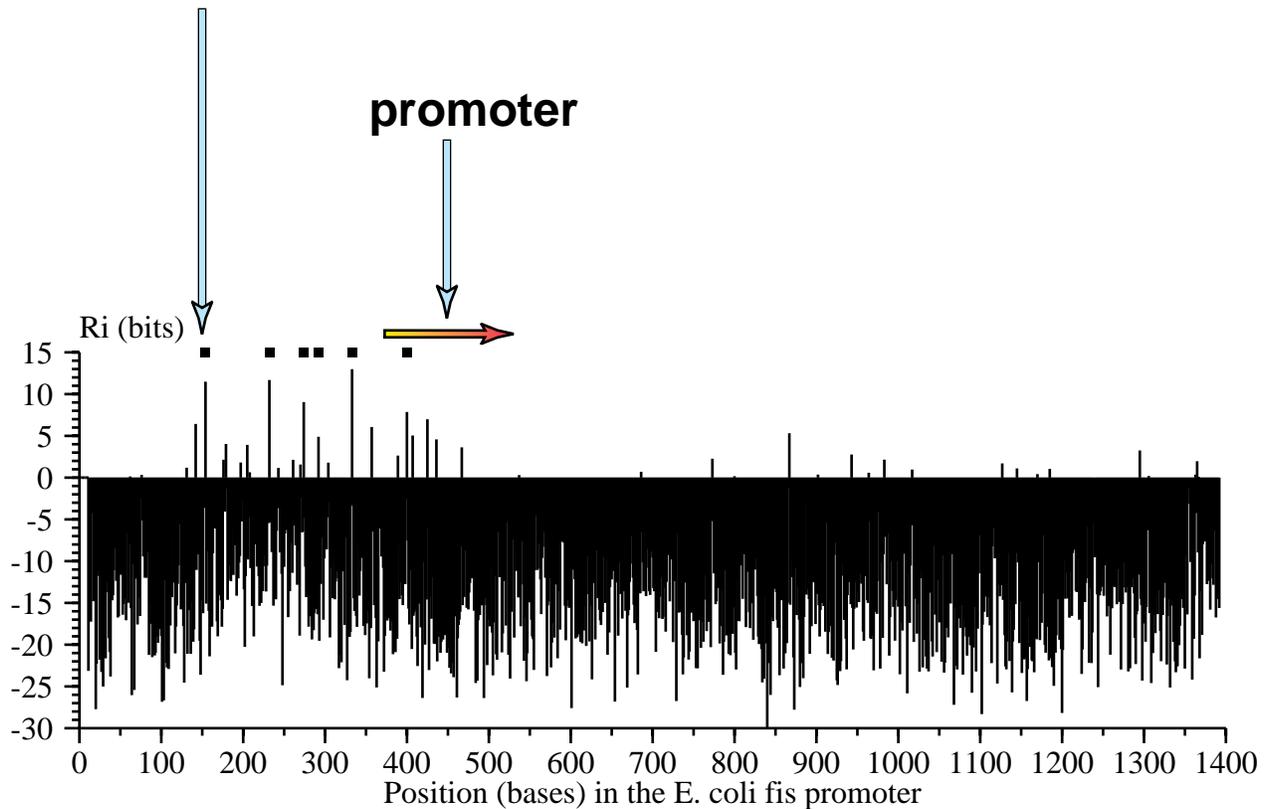
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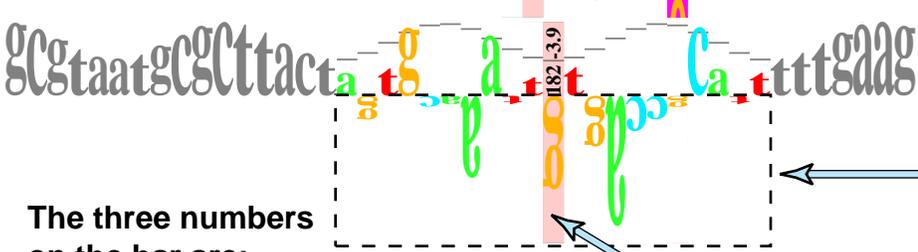
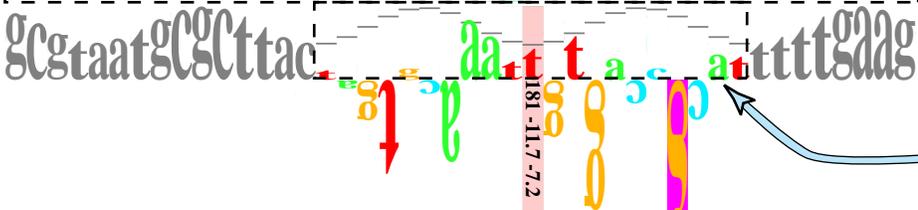
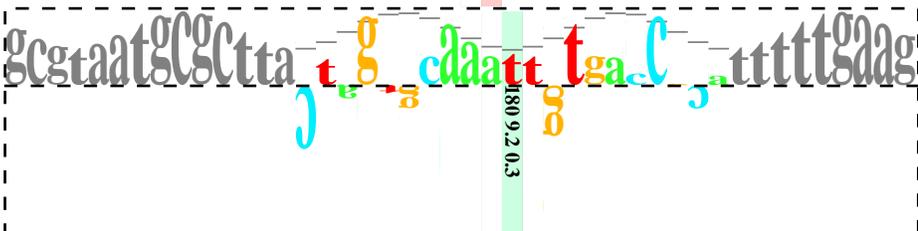
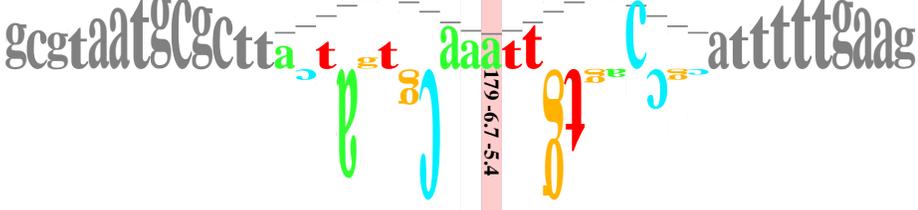
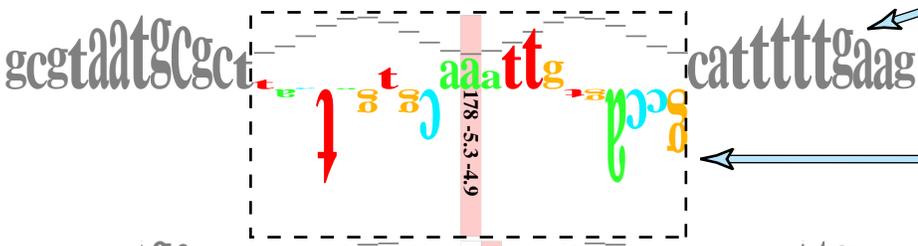


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❄️ WALKER

How well do the bases of a sequence match to functional binding sites?

5 copies of a single sequence are below:



The cosine wave represents the orientation of the DNA facing the protein.

The Walker is the colored letters.

The weight matrix is for the *E. coli* Fis protein.

The vertical bar is a scale: the top is at 2 bits the middle is at 0 bits the bottom is at -4 bits

Letters that go up are preferred at that position.

Letters that go down are detrimental at that position (purple goes below -4 bits).

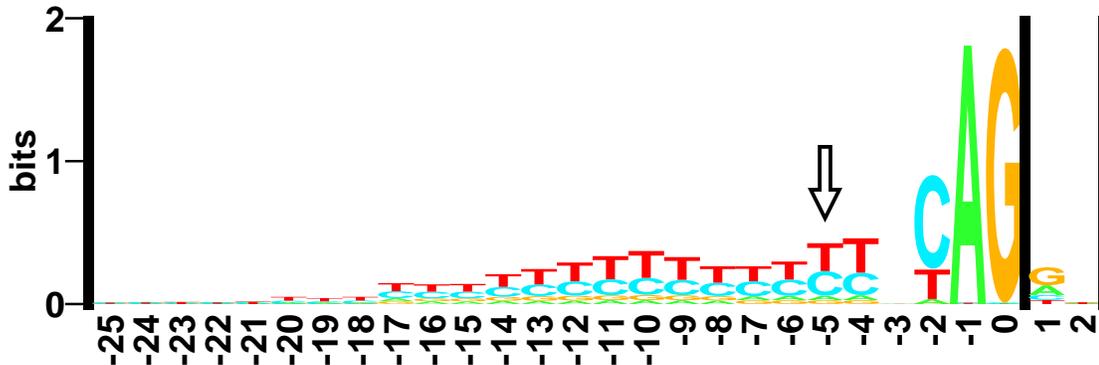
The three numbers on the bar are:

- * position
- * R individual
- * standard deviations from mean (Z)

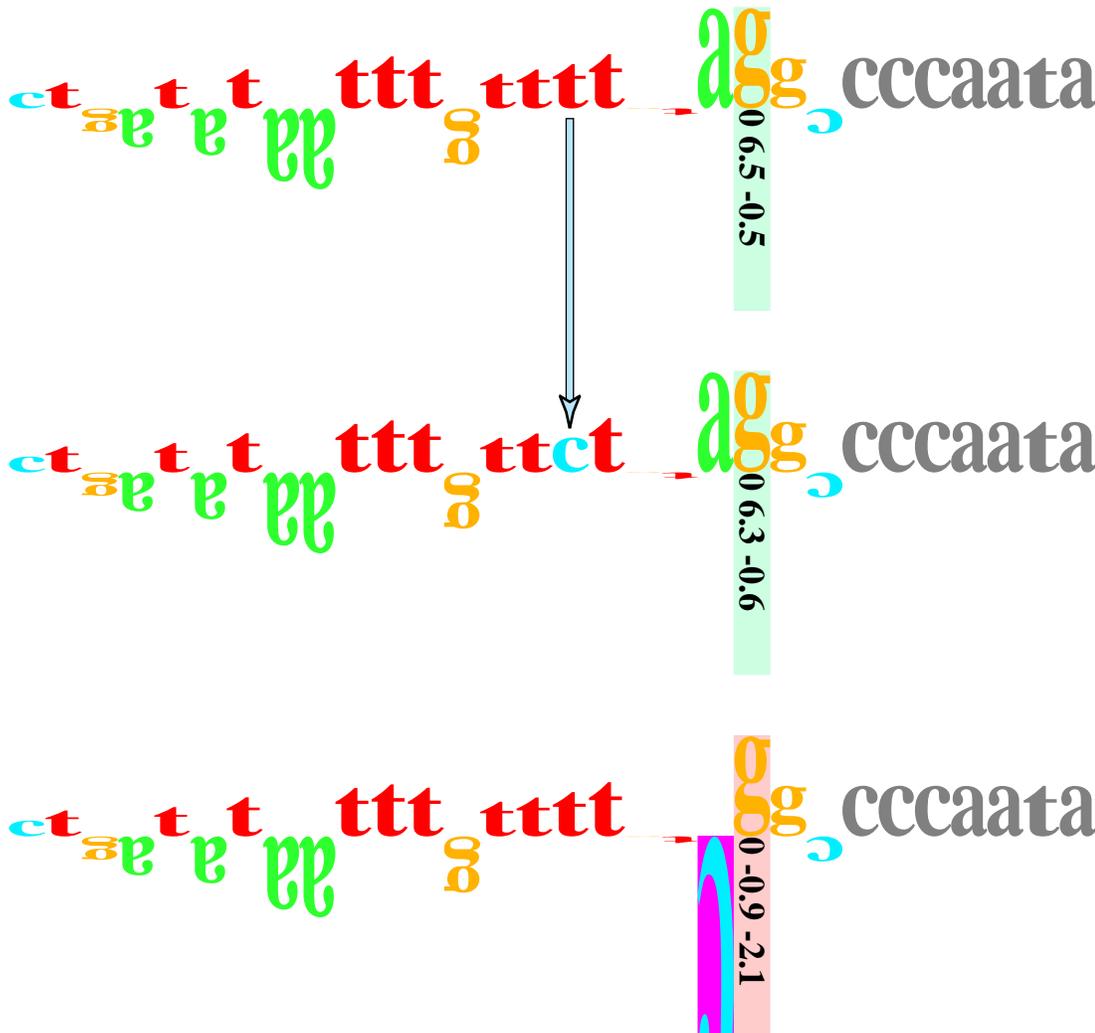
A green vertical bar indicates a binding site, red one means it is probably not a site.

Is a sequence change a Mutation or a Polymorphism?

A T to C change seen in a splice acceptor of hMSH2 was interpreted to be the mutation which causes familial nonpolyposis colon cancer (Fishel *et al.*, Cell 75:1027-1038, 1993):



The sequence logo shows nearly equal frequencies of bases there.



wild-type as seen by a walker

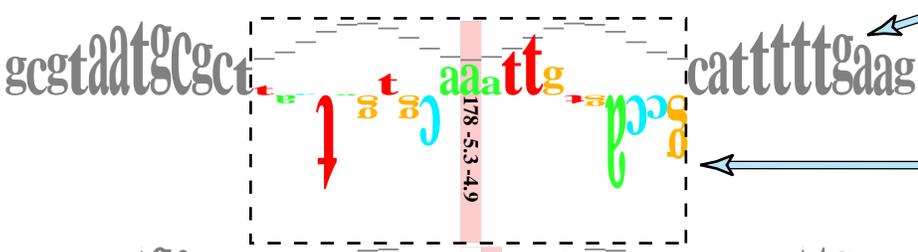
The walker shows it is a polymorphism.

This is what a strong mutation would look like.

WALKER

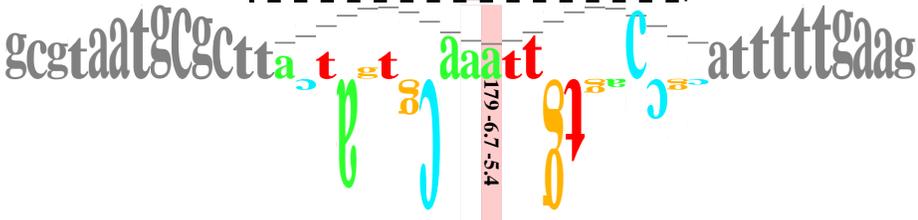
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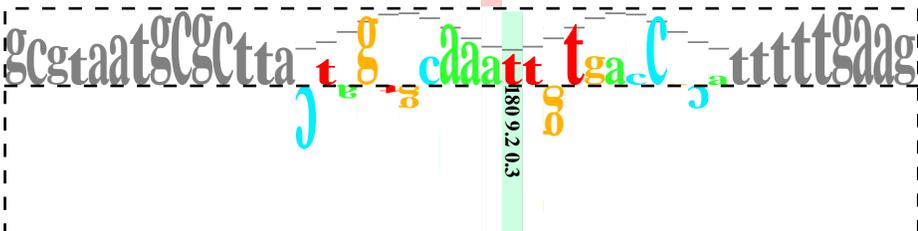


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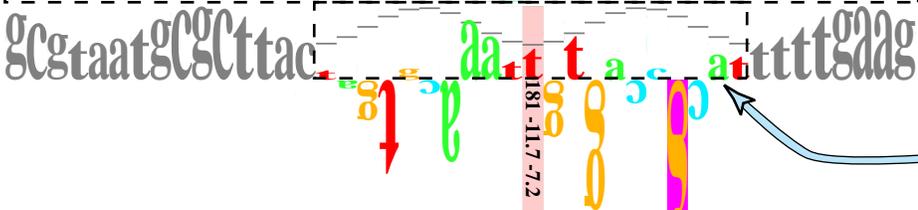
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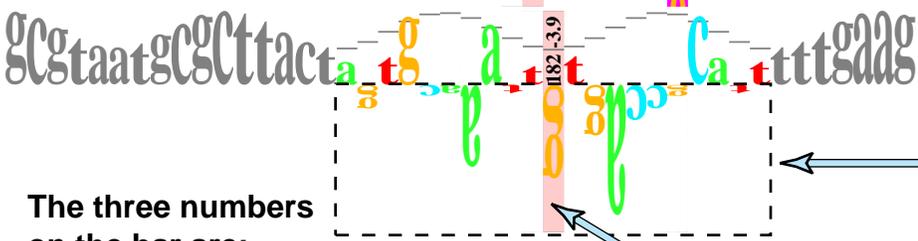
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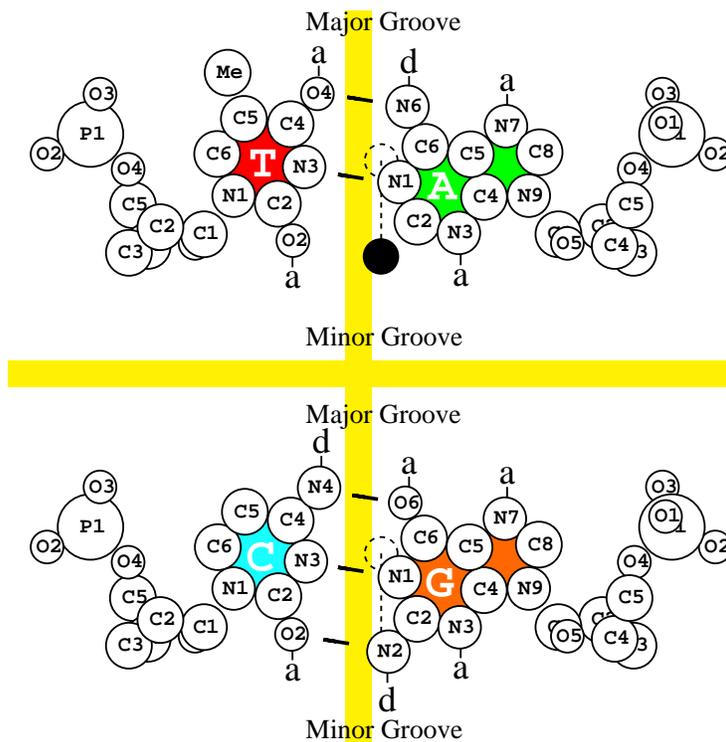
INFORMATION THEORY

A **BIT** measures the choice between 2 equally likely possibilities:



one bit
is like a knife
slice

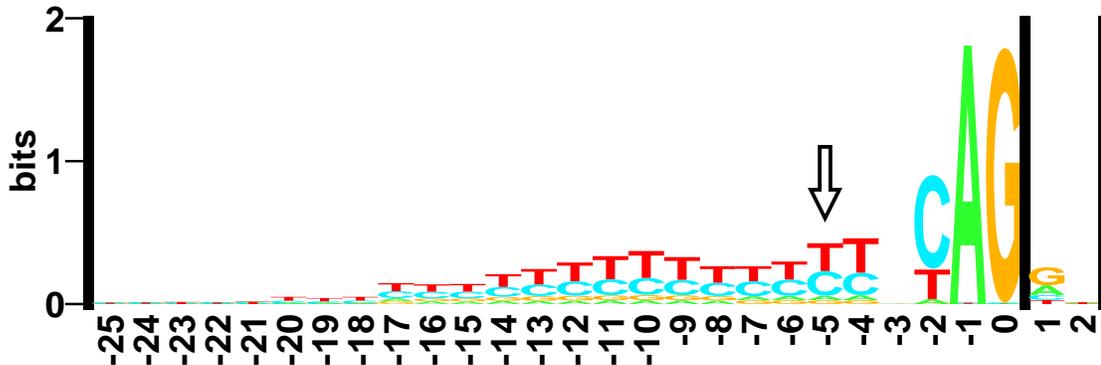
To choose one base in 4
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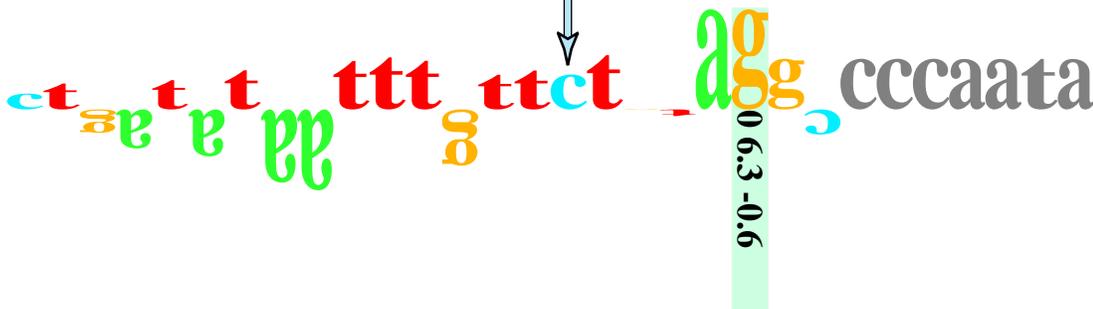
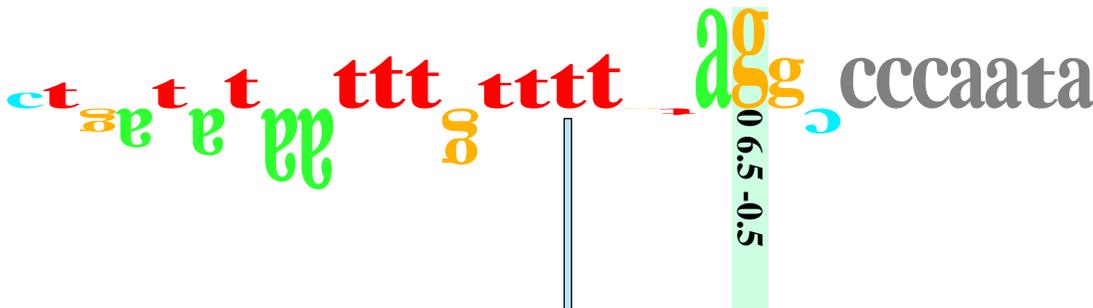
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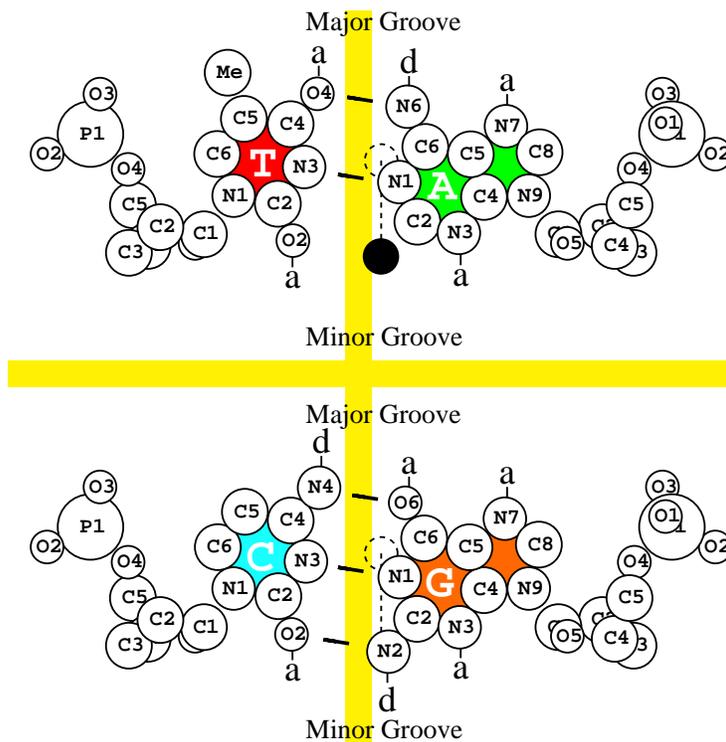
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Information measured in bits

❄ is additive

❄ is well supported mathematically

❄ measures sequence conservation

❄ is related to the entropy

❄ is calculated as a decrease in the uncertainty H:

$$H = -\sum p_i \log_2 p_i \text{ (bits)}$$

as

$$R = -\Delta H \quad \text{(bits/symbol)}$$

R is the Rate of information transmission.

Every sequence has an individual information

	-----	+++++	
	9876543210	123456789	
		
1	gtatcaccgcccagtggtat	17.7	bits
2	ataccactggcggtgatac	17.7	bits
3	tcaacaccgcccagagataa	19.3	bits
4	ttatctctggcggtgttga	19.3	bits
5	ttatcaccgcagatggtta	15.7	bits
6	taaccatctgcggtgataa	15.7	bits
7	ctatcaccgcaaggataa	17.3	bits
8	ttatcccttgcggtgatag	17.3	bits
9	ctaacaccgtgcggtgga	11.0	bits
10	tcaacacgcacgggtgtag	11.0	bits
11	ttacctctggcggtgataa	21.5	bits
12	ttatcaccgcccagaggtaa	21.5	bits

$$\Sigma = 205 \text{ bits}$$

$$\Sigma / 12 = 17.1 \text{ bits}$$

The average of the individual information values for the original sequence is the same as the sequence conservation and the area under the sequence logo.

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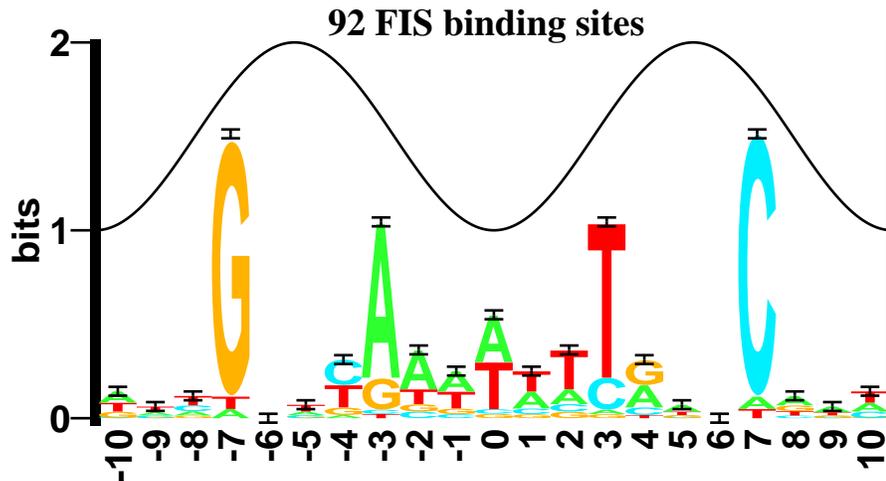
	-----	+++++++	
	9876543210	123456789	
		
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4	ttatctctggcgggtgattga	19.3	bits
5	ttatcaccgcccagatggtta	15.7	bits
6	taaccatctgcggtgataa	15.7	bits
7	ctatcaccgccaaggataa	17.3	bits
8	ttatcccttgccggatag	17.3	bits
9	ctaacaccgtgctgttga	11.0	bits
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Individual Information



The area under a sequence logo is the total sequence conservation.

The mathematics makes it look like an average.

QUESTION: What is it the average of?

ANSWER: The information of each individual binding site.

This is found from the individual information weight matrix as:

$$R_i(b,l) = 2 + \log_2 f(b,l)$$

where $f(b,l)$ is the frequency of each base b at every position l in the binding site.

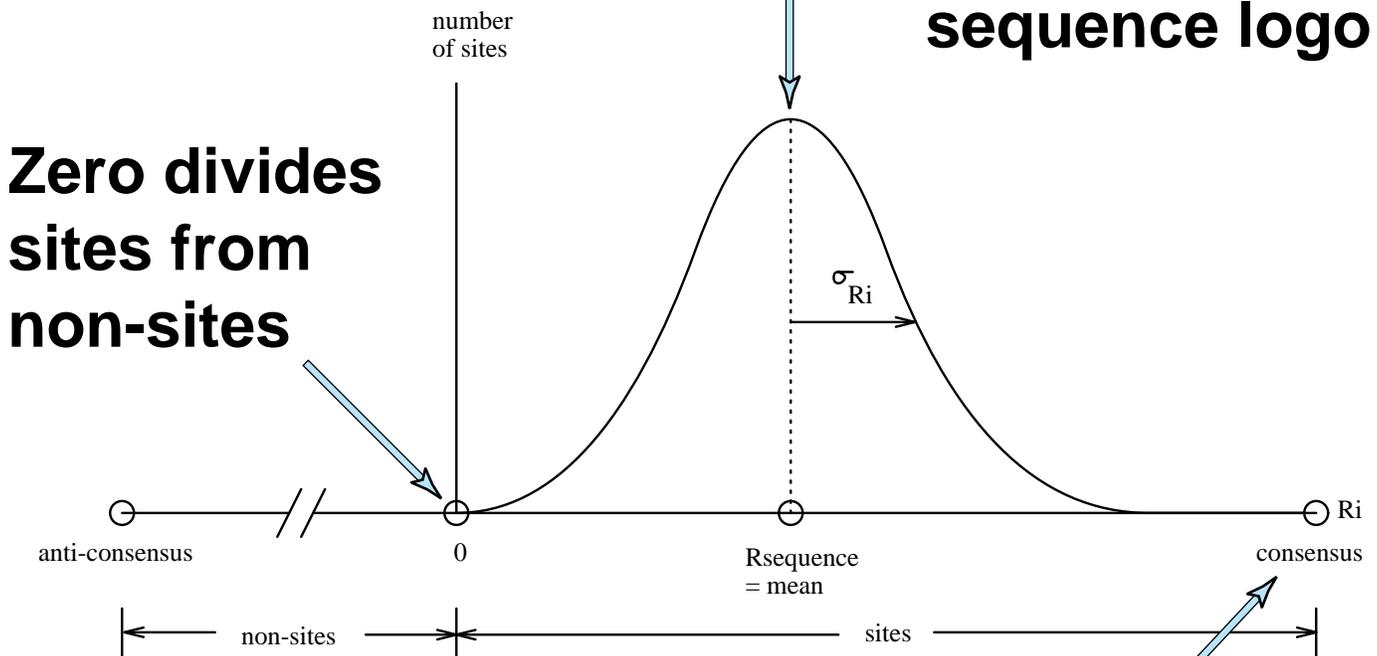
number of
bases b
at each
position l

l	$n(a,l)$	$n(c,l)$	$n(g,l)$	$n(t,l)$	$R_i(a,l)$	$R_i(c,l)$	$R_i(g,l)$	$R_i(t,l)$
-10	36	7	21	28	0.622843	-1.739727	-0.154765	0.260273
-9	20	19	15	38	-0.225154	-0.299154	-0.640191	0.700846
-8	18	24	11	39	-0.377157	0.037881	-1.087650	0.738320
-7	3	0	85	4	-2.962119	-6.539159	1.862309	-2.547082
-6	24	19	29	20	0.037881	-0.299154	0.310899	-0.225154
-5	20	18	15	39	-0.225154	-0.377157	-0.640191	0.738320
-4	5	40	12	35	-2.225154	0.774846	-0.962119	0.582201
-3	73	2	15	2	1.642743	-3.547082	-0.640191	-3.547082
-2	53	9	10	20	1.180838	-1.377157	-1.225154	-0.225154
-1	40	8	11	33	0.774846	-1.547082	-1.087650	0.497312
0	42	4	4	42	0.845235	-2.547082	-2.547082	0.845235
1	33	11	8	40	0.497312	-1.087650	-1.547082	0.774846
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3	2	15	2	73	-3.547082	-0.640191	-3.547082	1.642743
4	35	12	40	5	0.582201	-0.962119	0.774846	-2.225154
5	39	15	18	20	0.738320	-0.640191	-0.377157	-0.225154
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$R_i(b,l)$ weight matrix

Individual Information Distributions

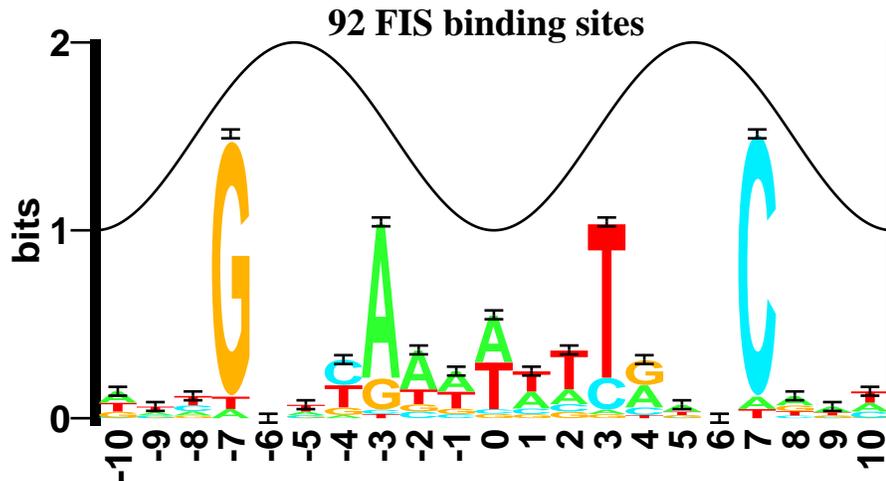
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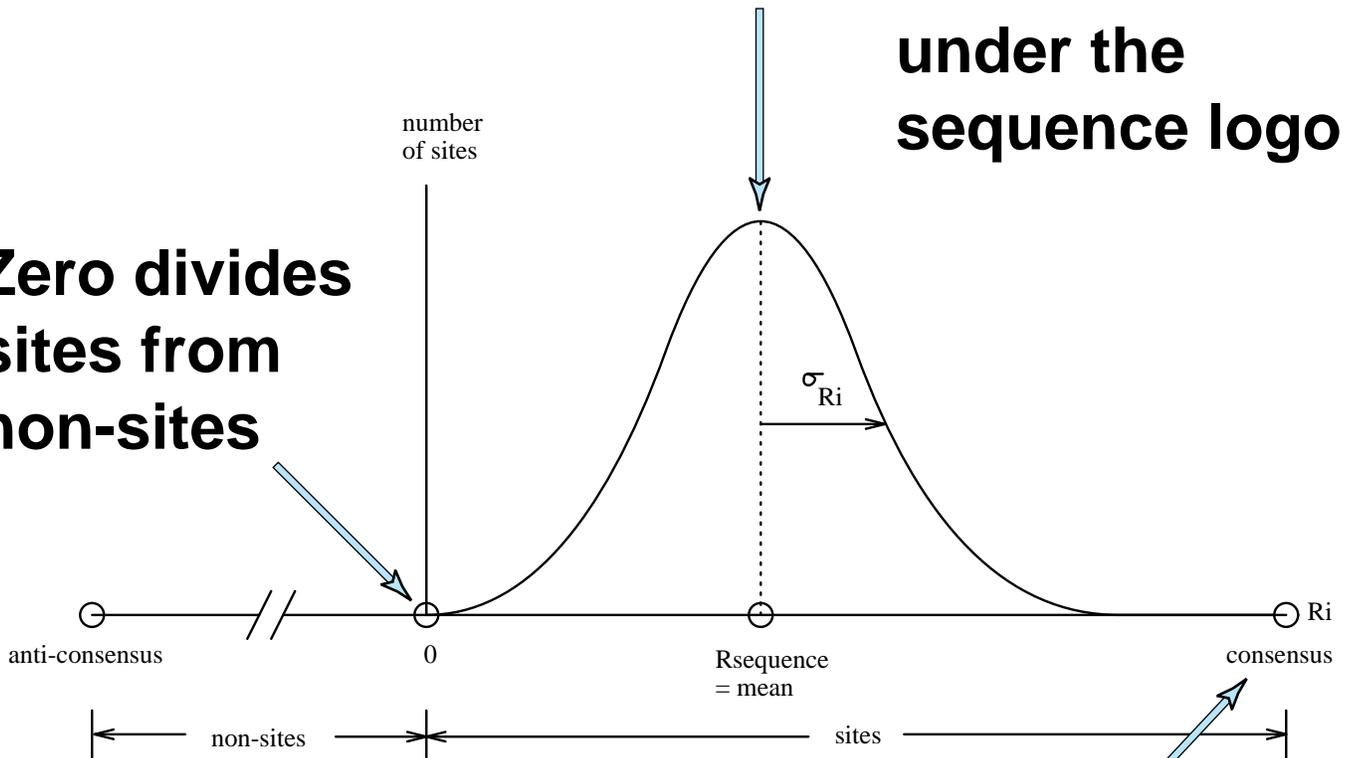
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